

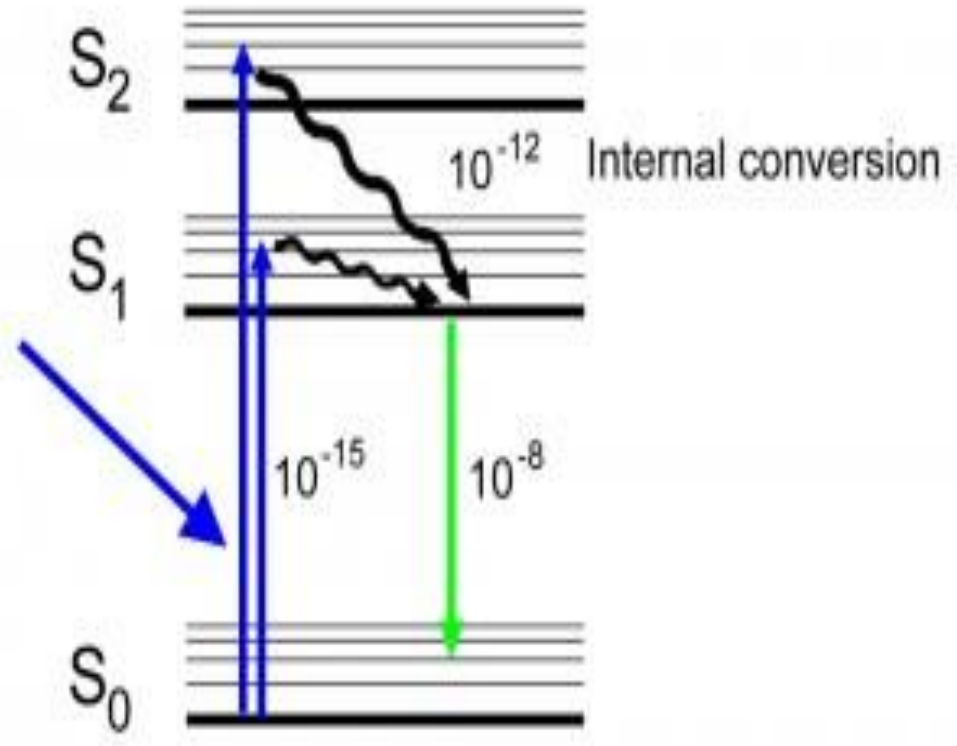
Fluorescence Spectroscopy



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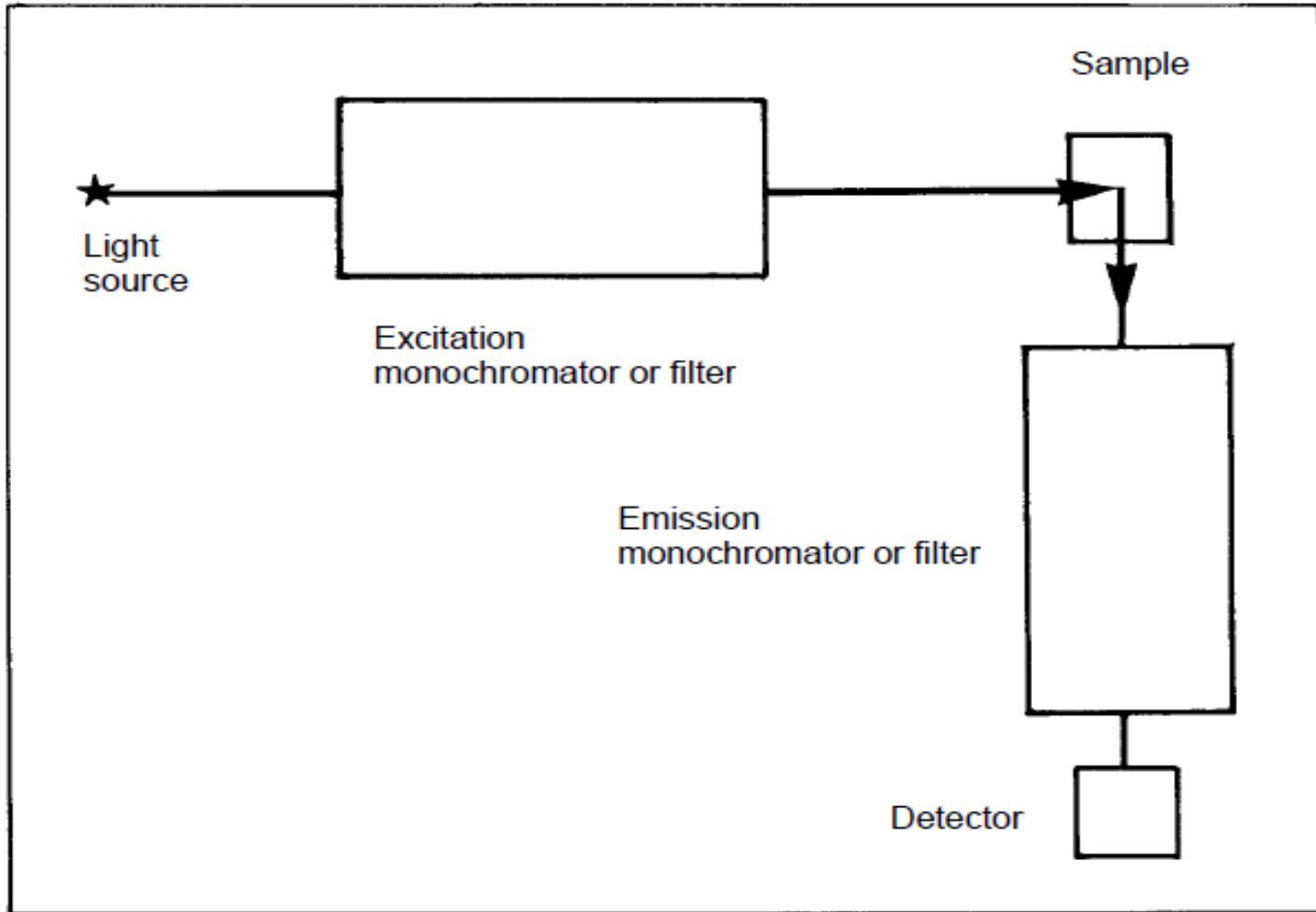
Fluorescence

- Electron in the ground state is excited to a higher energy state
- After loss of some energy in vibrational relaxation, the high energy electron returns back to the ground state by emitting fluorescent photon.



[7]

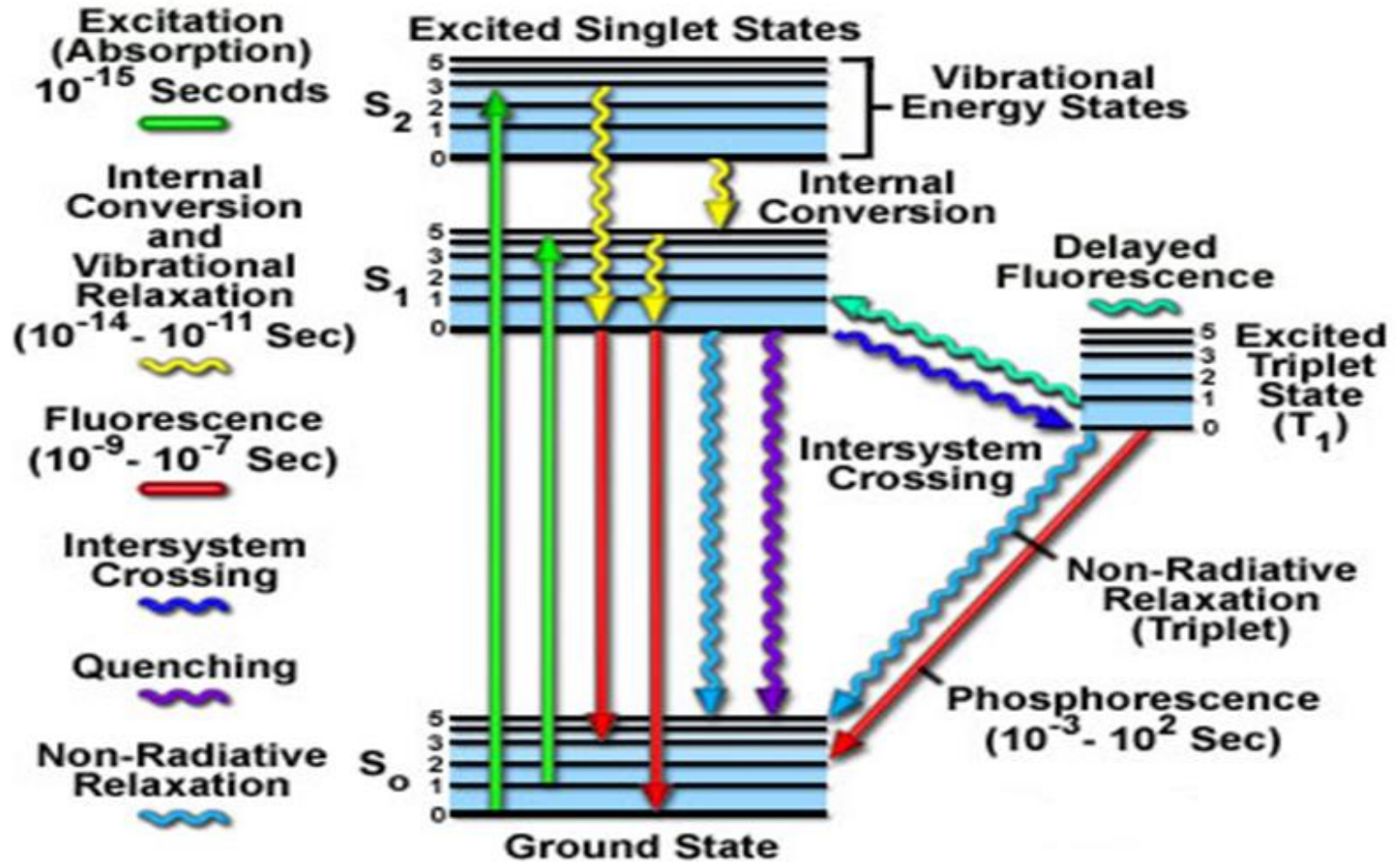
Experimental Set-up



[7]

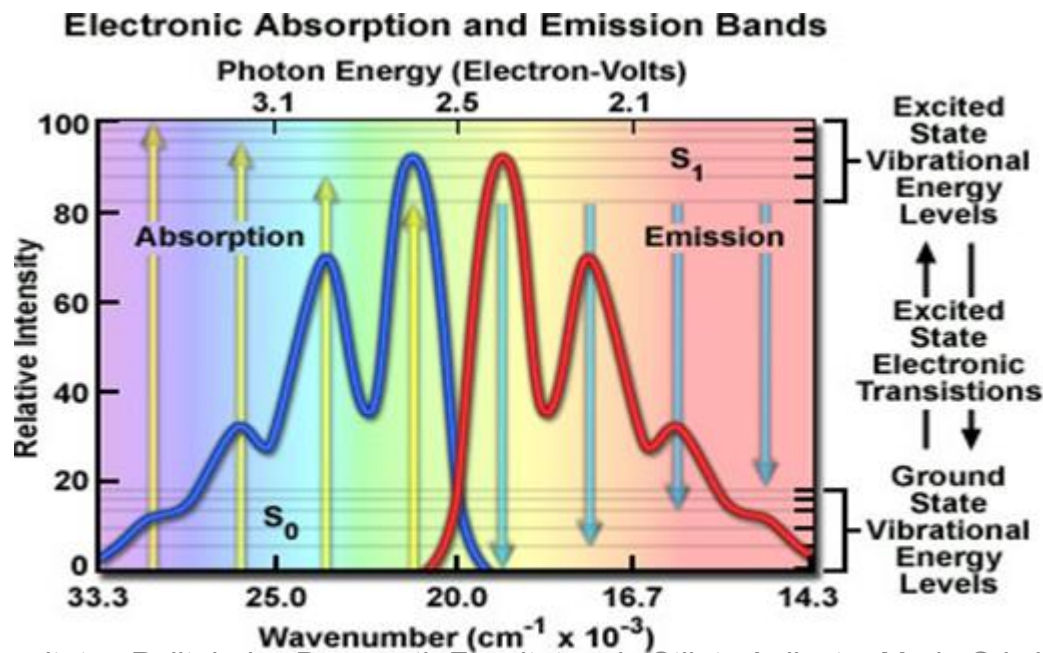
- Molecule that absorbs light = chromophor
- Molecule that emits light= fluorophor
- Fluorophor
 - Intrinsic fluorophores: aromatic amino acids
 - Extrinsic fluorophores :can be linked covalently or not to macromolecules such as peptides, proteins, membranes, or DNA.

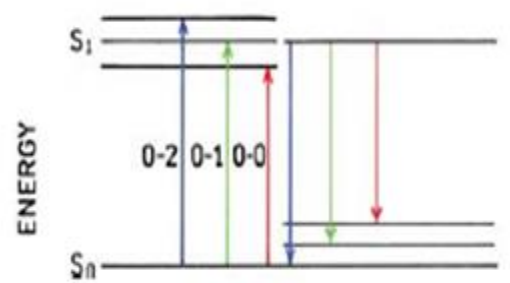
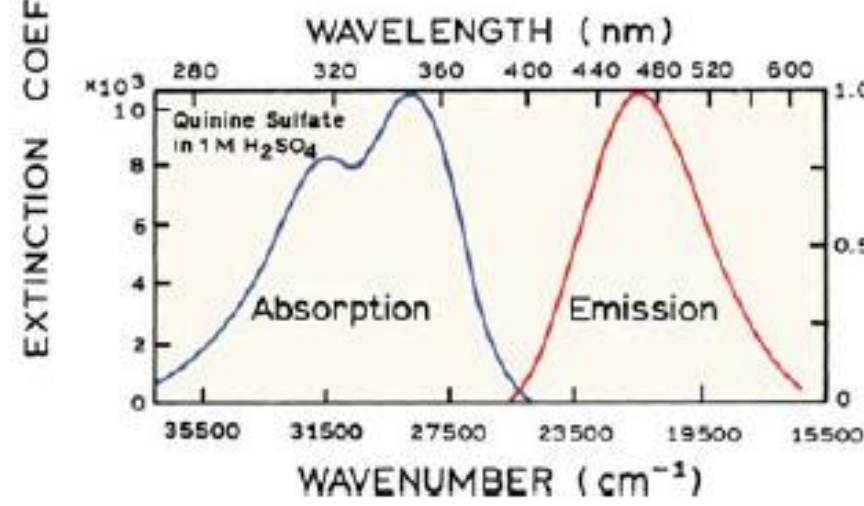
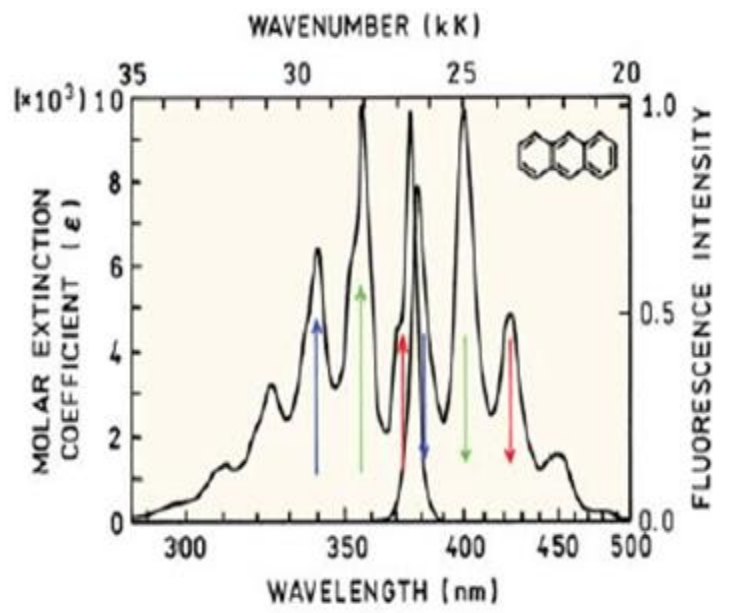
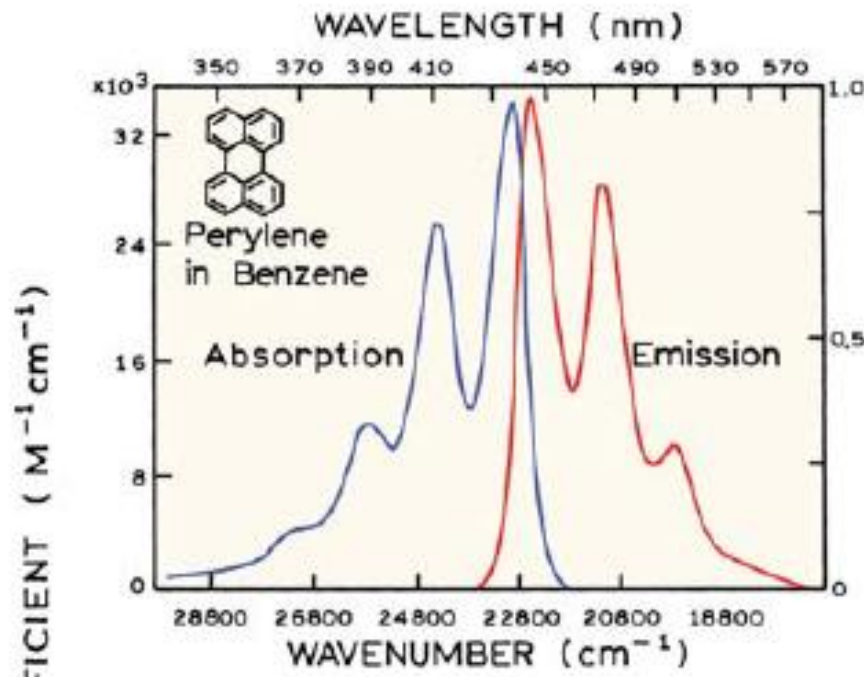
Jablonski Energy Diagram



[6]

- Stokes shift
- Intensity not dependent of the excitation λ
- Mirror rule





Mirror-image rule and Franck-Condon factors. The absorption and emission spectra are for anthracene. The numbers 0, and 2 refer to vibrational energy levels. From [1].

Quantum Yield. Fluorescence lifetime

- Quantum yield (Q) is the (dimensionless) ratio of photons emitted to the number of photons absorbed.

$$Q = \frac{\Gamma}{\Gamma + k_{nr}}$$

- Γ is fluorescence decay rate and k_{nr} is combined nonradiative decay rate

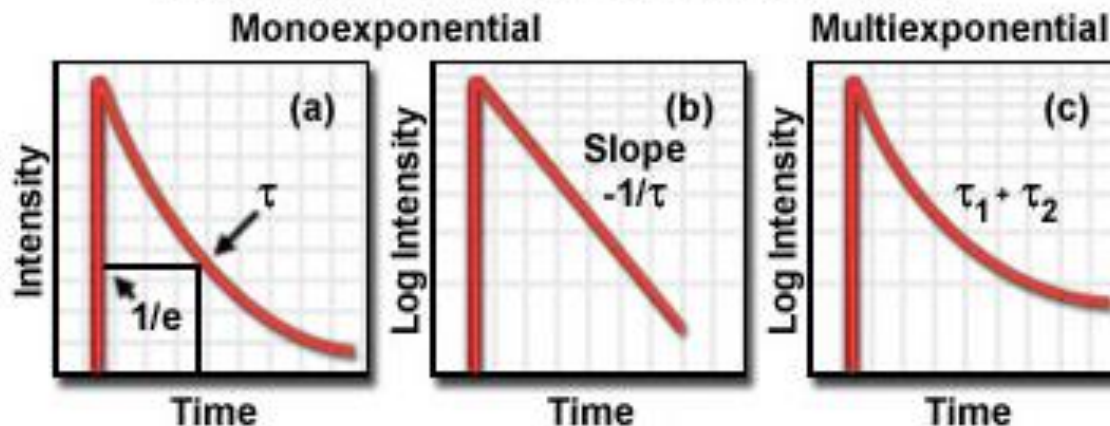
- Lifetime (τ) is an average value of time spent in the excited state
- The lifetime of the fluorophore in the absence of nonradiative processes is called the intrinsic or natural lifetime, τ_n

$$\tau = \frac{1}{\Gamma + k_{nr}} \quad \tau_n = \frac{1}{\Gamma} \quad \tau_n = \tau / Q$$

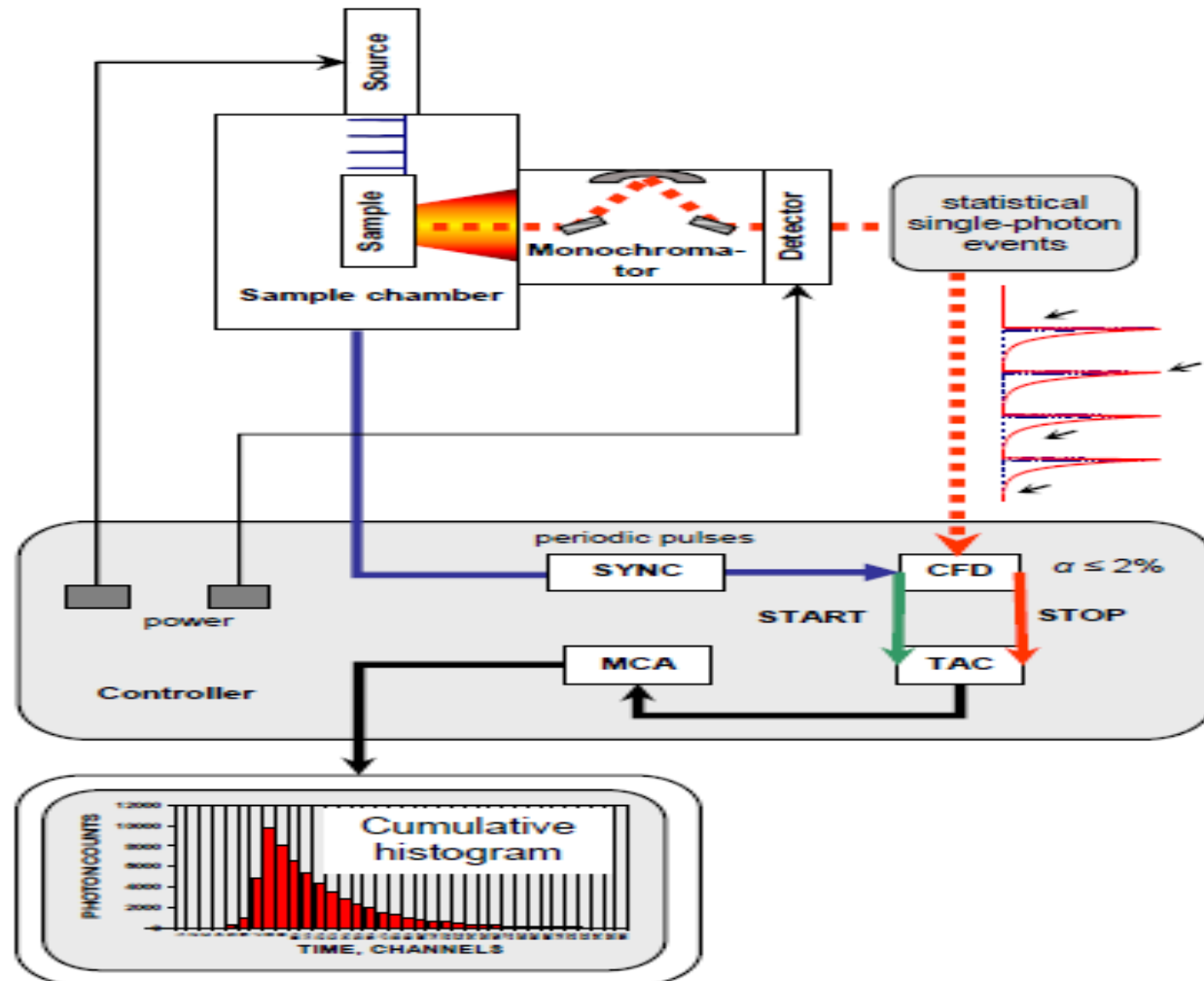
- The fluorescence lifetime is defined as the time in which the initial fluorescence intensity of a fluorophore decays to $1/e$ of the initial intensity

$$I(t) = I_0 e^{-\frac{t}{\tau}}$$

Fluorescence Lifetime Decay Profiles



Time-Correlated Single-Photon Counting (TCSPC) measurements



Quenching

- External molecules added to the fluorescent system can quench fluorescence intensity and therefore quantum yield
- Dynamic quenching: quenchers will decrease the fluorescence while entering in collision with the fluorophore
- Static quenching: quenchers form a nonfluorescent complex with the fluorophore

$$I/I_0 = 1 + K_{SV}[Q] = 1 + K_q\tau_0$$

- I_0 and I - fluorescence intensities observed in the absence and presence, respectively, of quencher
- $[Q]$ - the quencher concentration
- K_{SV} - the Stern-Volmer quenching constant
- K_q -bimolecular quenching rate constant
- τ_0 -excited state lifetime in the absence of quencher

FRET- Förster energy transfer

- Fret or energy transfer at a distance occurs between two molecules, a donor (the excited fluorophore), and an acceptor (a chromophore or fluorophore).
- Energy is transferred by resonance.

FRET

k_T - rate of energy transfer

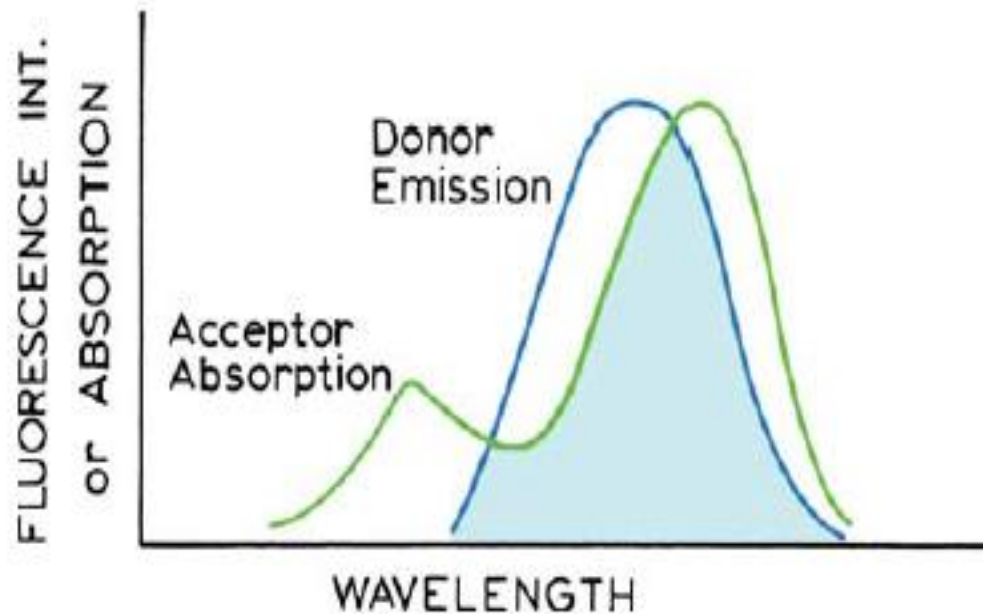
R_0 - Förster distance

r -distance between D and A

τ_D - lifetime of the donor in the absence of energy transfer



$$k_T = \frac{1}{\tau_D} \left(\frac{R_0}{r} \right)^6$$

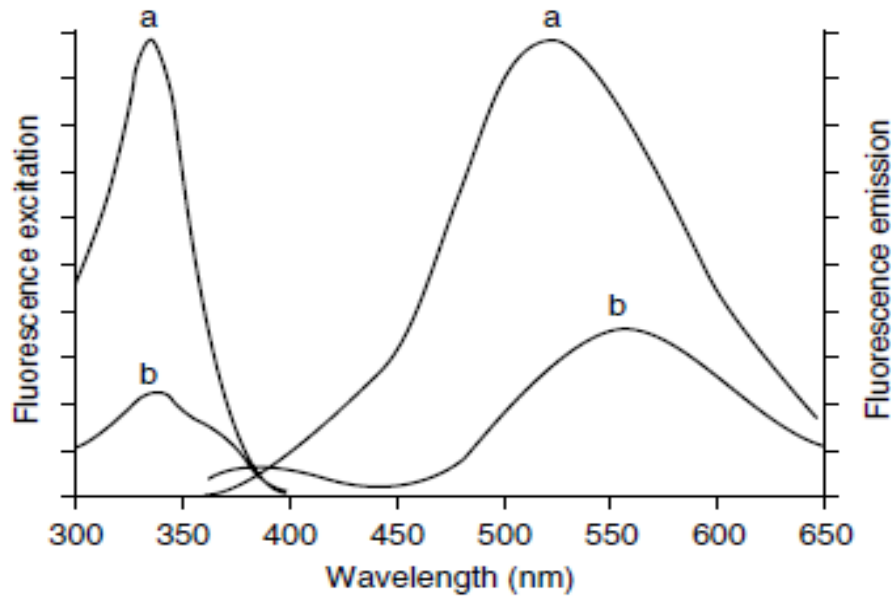


Lackowicz, J. R., Principles of Fluorescence Spectroscopy, Springer, 2006

Applications

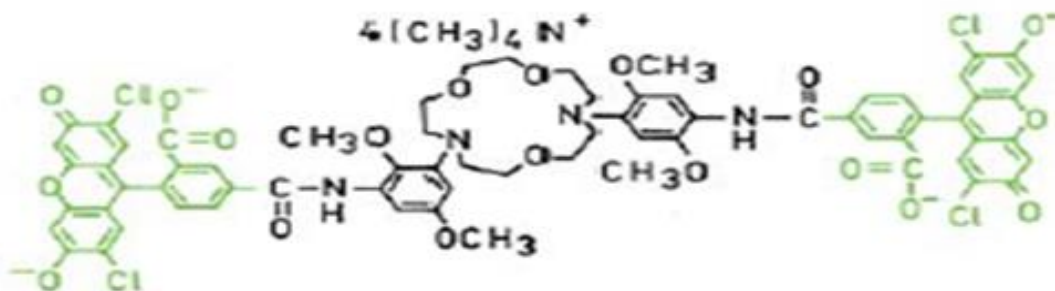
- Biology
- Ion detectors
- Medicine
- Nanoscience
- Molecules labeling
- DNA

Ion detectors

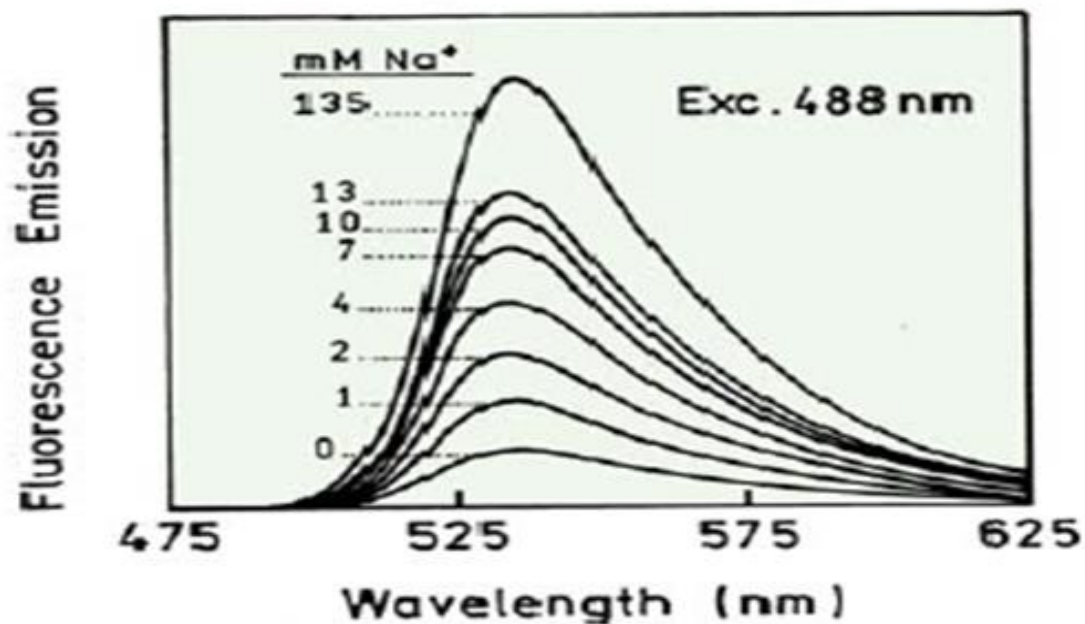


Fluorescence excitation (detected at 505 nm) and emission (excited at 340 nm) spectra of SBFI in pH 7.0 buffer containing 135 mM (a) or zero (b) Na^+ .

[2]



a c

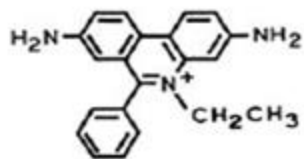


Effects of sodium on the emission of Sodium Green.

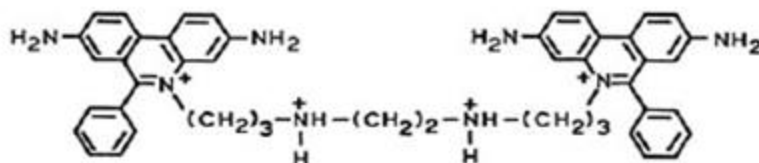
Lackowicz, J. R., Principles of Fluorescence Spectroscopy, Springer, 2006

DNA

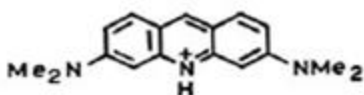
- Deoxyribonucleic acid is weakly or nonfluorescent
- Several dyes bind spontaneously to DNA—such as acridines, ethidium bromide, and other planar cationic species
- Staining of cells with dyes that bind to DNA is used to visualize and identify chromosomes



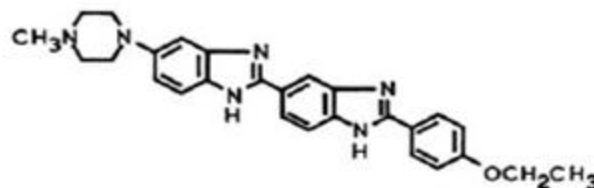
Ethidium Bromide
518/605 nm



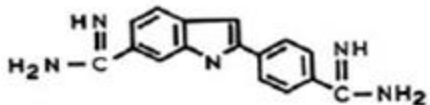
Ethidium Homodimer
528/617 nm



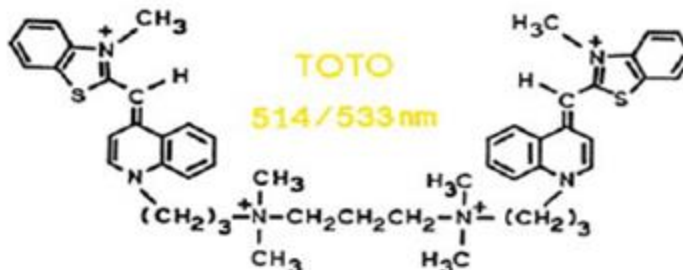
Acridine Orange
500/526 nm DNA
460/650 nm RNA



Hoechst 33342
350/460 nm



DAPI
355/461 nm



TOTO
514/533 nm

Representative DNA probes. Excitation and emission wavelengths refer to DNA-bound dye.

Conclusions

- Fluorescence spectroscopy
- Lifetime measurements
- Concentration measurements
- Multiple applications

References

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2. Wang, X. F., Herman, B., Fluorescence lifetime spectroscopy and imaging. Principles and applications in biomedical diagnostics, CRC Press, Taylor and Francis Group, 2015
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